

## Freeform Search

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US Pre-Grant Publication Full-Text Database
US Patents Full-Text Database
US OCR Full-Text Database
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JPO Abstracts Database
Derwent World Patents Index
IBM Technical Disclosure Bulletins

Term: L9 and (methyl or alkyl or alkyloxy or aryl) 

Display: 10 Documents in Display Format: [-] Starting with Number 1

Generate:  Hit List  Hit Count  Side by Side  Image

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**Search** **Clear** **Interrupt**

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### Search History

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DATE: Wednesday, April 21, 2004 [Printable Copy](#) [Create Case](#)

<u>Set</u>	<u>Hit</u>	<u>Set</u>
<u>Name</u>	<u>Count</u>	<u>Name</u>
side by side		result set
DB=USPT,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ		
<u>L10</u> L9 and (methyl or alkyl or alkyloxy or aryl)	6	<u>L10</u>
<u>L9</u> (primer\$1 or oligonucleotide\$1) near5 no\$1 complementary near5 modif\$7	10	<u>L9</u>
<u>L8</u> L7 and 2 end and 3 end	5	<u>L8</u>
<u>L7</u> L6 and DNA polymerase\$1	38	<u>L7</u>
<u>L6</u> L5 and (alkyl or alkyloxy or alkylamino or aryl or aryloxy)	44	<u>L6</u>
<u>L5</u> L4 and modif\$7 nucleotide\$1	134	<u>L5</u>
<u>L4</u> (primer\$1 or oligonucleotide\$1 or probe\$1) near5 no\$1 complementary	1079	<u>L4</u>
<u>L3</u> l1 and ((no\$1 or less) near5 exten\$5)	6	<u>L3</u>
<u>L2</u> L1 and exten\$5	6	<u>L2</u>
<u>L1</u> (primer\$1 or oligonucleotide\$1 or probe\$1 or nucleic acid sequence\$1) near5 no\$1 complementary near5 modif\$3 near5 nucleotide\$1	6	<u>L1</u>

END OF SEARCH HISTORY



origin of affinity enhancement. The following general factors are important. The loop constraint, or closure of the 2 anticodon sequences into hairpin loops, accounts for about a factor 50 in the affinity. Dangling ends, or **non-complementary** nucleotides at the end of the double helix contribute strongly to the affinity. **Modified nucleotides**, like the Y base, in the dangling ends can contribute a special stabilization of up to a factor of 7. These observations can be understood in terms of a model in which the short 3 base-pair helix is sandwiched between stacked bases and hence stabilized. The potential importance of loop-loop interactions and stacking effects for codon-anticodon bonding is emphasized. A possible simple physical basis may exist for the evolutionary choice of a triplet coding system.

AB. . . complementary trinucleotides. The association rate constant (3 + 106 M-1 at 25° C) is similar to typical values observed for **oligonucleotides**, so the enhanced affinity in the tRNA · tRNA complex is due entirely to a much slower dissociation than expected. . . of the 2 anticodon sequences into hairpin loops, accounts for about a factor 50 in the affinity. Dangling ends, or **non-complementary** nucleotides at the end of the double helix contribute strongly to the affinity. **Modified nucleotides**, like the Y base, in the dangling ends can contribute a special stabilization of up to a factor of 7.. . .

=> s (primer# or oligonucleotide#) (P) (no# complementary or no# hybridiz#####) (p) modif##### nucleotide#  
L3 1 (PRIMER# OR OLIGONUCLEOTIDE#) (P) (NO# COMPLEMENTARY OR NO# HYBRID IZ#####) (P) MODIF##### NUCLEOTIDE#

=> d 13 bib ab kwic

L3 ANSWER 1 OF 1 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN  
AN 1976:212966 BIOSIS  
DN PREV197662042966; BA62:42966  
TI STUDIES OF THE COMPLEX BETWEEN TRANSFER RNA WITH COMPLEMENTARY ANTI CODONS PART 1 ORIGINS OF ENHANCED AFFINITY BETWEEN COMPLEMENTARY TRIPLETS.  
AU GROSJEAN H; SOLL D G; CROTHERS D M  
SO Journal of Molecular Biology, (1976) Vol. 103, No. 3, pp. 499-519.  
CODEN: JMOBAK. ISSN: 0022-2836.  
DT Article  
FS BA  
LA Unavailable  
AB The temperature-jump method was used to study the complex between yeast tRNAPhe and Escherichia coli tRNAGlu, which have the complementary anticodons GmAA and s2UUC, respectively. The binding constant (3.6 + 105 M-1 at 25° C) is about 6 orders of magnitude larger than expected for 2 complementary trinucleotides. The association rate constant (3 + 106 M-1 at 25° C) is similar to typical values observed for **oligonucleotides**, so the enhanced affinity in the tRNA · tRNA complex is due entirely to a much slower dissociation than expected for a 3 base-pair helix. An association enthalpy of -25 kcal/mol, nearly twice as large as expected for 2 stacking interactions in a 3 base-pair helix was found. The association entropy (-58 cal/deg per mol) is close to the expected value. The reaction occurs with a single relaxation and therefore does not involve any slow reorganization of the tRNA molecule. Structural variations were studied to investigate the origin of affinity enhancement. The following general factors are important. The loop constraint, or closure of the 2 anticodon sequences into hairpin loops, accounts for about a factor 50 in the affinity. Dangling ends, or **non-complementary** nucleotides at the end of the double helix contribute strongly to the affinity. **Modified nucleotides**, like the Y base, in the dangling ends can contribute a special stabilization of up to a factor of 7. These observations can be understood in terms of a model in which the short 3

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=> s (primer# or oligonucleotide#) (p)modif##### nucleotide#  
L4 444 (PRIMER# OR OLIGONUCLEOTIDE#) (P) MODIF##### NUCLEOTIDE#

=> s 14 and (no# complementary or no# hybridiz#####)  
L5 5 L4 AND (NO# COMPLEMENTARY OR NO# HYBRIDIZ#####)

=> dup rem 15  
PROCESSING COMPLETED FOR L5  
L6 2 DUP REM L5 (3 DUPLICATES REMOVED)

=> d 16 1-2 bib ab kwic

L6 ANSWER 1 OF 2 MEDLINE on STN DUPLICATE 1  
AN 96428850 MEDLINE  
DN PubMed ID: 8831952  
TI Recognition of the **primers** containing different **modified nucleotide** units by the Klenow fragment of DNA polymerase I from **E coli**.  
AU Kolocheva T I; Levina A S; Nevinsky G A  
CS Novosibirsk Institute of Bioorganic Chemistry, Siberian Division of the Russian Academy of Sciences, Russia.  
SO Biochimie, (1996) 78 (3) 201-3.  
Journal code: 1264604. ISSN: 0300-9084.  
CY France  
DT Journal; Article; (JOURNAL ARTICLE)  
LA English  
FS Priority Journals  
EM 199612  
ED Entered STN: 19970128  
Last Updated on STN: 19970128  
Entered Medline: 19961203  
AB A comparison of Km values and maximal rates of extension (Vmax) for primers containing different modified bases or mismatches, and fully complementary primers of the same length catalyzed by the Klenow fragment of **E coli** DNA polymerase I was carried out. Base modifications include T-T dimers and apurinic sites. In the case of mismatch, the number of complementary bases from the 3'-terminus to the **non-complementary** nucleotide determines the efficiency of substrate incorporation, which is a measure of degree of interaction of the enzyme with its primer template. Differently, removal of one base in any position from the 3'-terminus of the primer is equivalent to shortening of the primer by one nucleotide unit, and decreases the affinity to the enzyme by 1.8-fold. Since apurinic sites fail to interfere with the efficiency of DNA synthesis, we suppose that the Klenow fragment of **E coli** DNA polymerase I does not participate in the correction of DNAs containing apurinic nucleotides units. Finally, the efficiency of elongation of the d(p primer was shown to decrease with an increase in T-T dimers in the primer. When the d(pT)10m primer contains about 2.6 T-T dimers per

TI molecule, the efficiency of its elongation decreases by a factor of 8-18. Recognition of the **primers** containing different **modified nucleotide** units by the Klenow fragment of DNA polymerase I from *E coli*.

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L6 ANSWER 2 OF 2 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN  
AN 1976:212966 BIOSIS  
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TI STUDIES OF THE COMPLEX BETWEEN TRANSFER RNA WITH COMPLEMENTARY ANTI CODONS PART 1 ORIGINS OF ENHANCED AFFINITY BETWEEN COMPLEMENTARY TRIPLETS.

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AB The temperature-jump method was used to study the complex between yeast tRNAPhe and *Escherichia coli* tRNAGlu, which have the complementary anticodons GmAA and s2UUC, respectively. The binding constant ( $3.6 \times 10^5$  M-1 at 25° C) is about 6 orders of magnitude larger than expected for 2 complementary trinucleotides. The association rate constant ( $3 \times 10^6$  M-1 at 25° C) is similar to typical values observed for **oligonucleotides**, so the enhanced affinity in the tRNA · tRNA complex is due entirely to a much slower dissociation than expected for a 3 base-pair helix. An association enthalpy of -25 kcal/mol, nearly twice as large as expected for 2 stacking interactions in a 3 base-pair helix was found. The association entropy (-58 cal/deg per mol) is close to the expected value. The reaction occurs with a single relaxation and therefore does not involve any slow reorganization of the tRNA molecule. Structural variations were studied to investigate the origin of affinity enhancement. The following general factors are important. The loop constraint, or closure of the 2 anticodon sequences into hairpin loops, accounts for about a factor 50 in the affinity. Dangling ends, or **non-complementary** nucleotides at the end of the double helix contribute strongly to the affinity. **Modified nucleotides**, like the Y base, in the dangling ends can contribute a special stabilization of up to a factor of 7. These observations can be understood in terms of a model in which the short 3 base-pair helix is sandwiched between stacked bases and hence stabilized. The potential importance of loop-loop interactions and stacking effects for codon-anticodon bonding is emphasized. A possible simple physical basis may exist for the evolutionary choice of a triplet coding system.

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